

# Package ‘RgnTX’

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**Title** Colocalization analysis of transcriptome elements in the presence of isoform heterogeneity and ambiguity

**Version** 1.0.0

**Description** RgnTX allows the integration of transcriptome annotations so as to model the complex alternative splicing patterns. It supports the testing of transcriptome elements without clear isoform association, which is often the real scenario due to technical limitations. It involves functions that do permutation test for evaluating association between features and transcriptome regions.

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**Author** Yue Wang [aut, cre],  
Jia Meng [aut]

**Maintainer** Yue Wang <yue.wang19@student.xjtlu.edu.cn>

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---

calculateShift	<i>Calculate positional shifting over transcriptome</i>
----------------	---

---

**Description**

The first step of calculating positional shift over transcriptome regions.

**Usage**

```
calculateShift(regions, disp, direction = "right", strand = "+")
```

**Arguments**

regions	A feature set, which should be a GRangesList object.
disp	A data frame object. It should have three columns, which are start: starting positions. Each value represents a starting position in each input feature; width: widths. Each value represents a width of each region to be picked from each feature; names: corresponding transcript ids.
direction	Either to be character "left" or "right", which means the direction to which the starting position is shifting. The former means moving to the direction of 5' while the latter means moving to 3'.
strand	Either to be "+" or "-".

**Value**

A GRanges object.

**See Also**

[extractRegions](#)

**Examples**

```
# Take five transcripts.
# Extract the last 200 nt regions from their CDS part.
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
trans.id.pstv <- c("170", "782", "974", "1364", "1387")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

# Download the CDS part of all transcriptome
cds.tx0 <- cdsBy(txdb, use.names = FALSE)

# pick the CDS part of these five transcripts
cds.p <- cds.tx0[trans.id.pstv]

width <- 200
disp.p.l <- data.frame(
  start = as.numeric(max(end(cds.p))),
  distance = width - 1,
  names = trans.id.pstv
)
R.p.l <- calculateShift(
  regions = cds.p, disp = disp.p.l,
  direction = "left", strand = "+"
)
```

distanceTx                      *Evaluation function*

---

### Description

Evaluation function. This function calculates the mean of the distance from each region of set RS1 to the closest region in RS2.

### Usage

```
distanceTx(A, B, beta = 0.2, ...)
```

### Arguments

A	Region set 1. A Granges or GRangesList object.
B	Region set 2. A Granges or GRangesList object.
beta	It is a user-defined argument that can filter out the corresponding percent of largest distance values. Default value is 0.2.
...	Any additional parameters needed.

### Value

A numeric object.

### See Also

[overlapWidthTx](#), [overlapCountsTx](#)

### Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
A <- randomizeTx(
  txdb, trans.ids,
  random_num = 20,
  random_length = 100
)
B <- randomizeTx(
  txdb, trans.ids,
  random_num = 20,
  random_length = 100
)
distanceTx(A, B, beta = 0.2)
```

---

extractRegions	<i>Extract regions</i>
----------------	------------------------

---

### Description

This function receives three arguments: the scope region set, the target region set and the type of strand. It returns a subset of target region set, which is the intersection of the target region set and the scope region set.

### Usage

```
extractRegions(regions_A, R, strand = "+")
```

### Arguments

regions_A	The scope region set. A GRangesList object. The name of each list element should be the transcript id that it pertains to.
R	The target region set. A GRanges object.
strand	The strand type of the transcripts. It has options "+" and "-".

### Value

A GRangesList object.

### See Also

[calculateShift](#)

### Examples

```
# Take five transcripts.
# Extract the last 200 nt regions from their CDS part.
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
trans.id.pstv <- c("170", "782", "974", "1364", "1387")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

# download the CDS part of all transcriptome
cds.tx0 <- cdsBy(txdb, use.names = FALSE)

# pick the CDS part of these five transcripts
cds.p <- cds.tx0[trans.id.pstv]

width <- 200
disp.p.l <- data.frame(
  start = as.numeric(max(end(cds.p))),
  distance = width - 1,
  names = trans.id.pstv
)
```

```
R.p.l <- calculateShift(regions = cds.p, disp = disp.p.l, direction = "left", strand = "+")
R.cds.last200 <- extractRegions(regions_A = cds.p, R = R.p.l, strand = "+")
```

---

```
getFormatCorrect      getFormatCorrect
```

---

**Description**

This function makes sure the two input region sets are in the correct format required by RgnTX evaluation functions.

**Usage**

```
getFormatCorrect(A, B)
```

**Arguments**

A	Region set 1. A Granges or GRangesList object.
B	Region set 2. A Granges or GRangesList object.

**Value**

A list object.

---

```
getPermSpaceByFeatures
                          Get permutation space for features
```

---

**Description**

This function returns a default permutation space for features with isoform ambiguity. The default permutation space of a feature is the aggregate of the multiple transcripts it may overlap with. It requires the input feature to be GRanges format.

**Usage**

```
getPermSpaceByFeatures(features, txdb, type = "mature")
```

**Arguments**

features	A GRanges object.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of regions over transcriptome.

**Value**

A list object, which contains two elements.

- perm.space: A GRangesList object that includes all the transcripts input features may overlap with.
- index: It contains a series of numbers indicating which feature these transcripts are respectively associated with.

**See Also**

[getPermSpaceByTxID](#), [getPermSpaceByType](#)

**Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
permSpace <- getPermSpaceByFeatures(features = m6A_sites_data[1:100], txdb)
```

---

getPermSpaceByTxID      *Get permutation space by specifying transcript ids*

---

**Description**

This function returns 5'UTR/CDS/3'UTR/mRNA/full part of transcriptome regions grouped by corresponding transcript ids.

**Usage**

```
getPermSpaceByTxID(trans_ids = "all", txdb, type = "mature")
```

**Arguments**

trans_ids	A character object. The transcript ids. Default is "all". If it takes the default value "all", the space that users get will be the whole transcriptome.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.

**Value**

A GRangesList object.

**See Also**

[getPermSpaceByType](#), [getPermSpaceByFeatures](#)

### Examples

```
trans.ids <- c("170", "782", "974", "1364", "1387")
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
permSpace <- getPermSpaceByTxID(trans.ids, txdb)
```

---

getPermSpaceByType      *Get permutation space by specifying type*

---

### Description

This function can return 5'UTR/CDS/3'UTR/mRNA/full part of transcriptome regions, following the format required by the main permutation test functions.

### Usage

```
getPermSpaceByType(txdb, type = "mature")
```

### Arguments

txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.

### Value

A GRangesList object.

### See Also

[getPermSpaceByTxID](#), [getPermSpaceByFeatures](#)

### Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
permSpace <- getPermSpaceByType(txdb, type = "CDS")
```



---

getPvalZscore	<i>getPvalZscore</i>
---------------	----------------------

---

**Description**

Calculate a p-value and z-score based on observed value and random evaluation values.

**Usage**

```
getPvalZscore(orig.ev, rand.ev, pval_z = FALSE)
```

**Arguments**

orig.ev	Observed value.
rand.ev	Random evaluation values.
pval_z	Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, the p-value is calculated based on a z-test.

**Value**

A p-value and a z-score.

---

getStopCodon	<i>getStopCodon</i>
--------------	---------------------

---

**Description**

Get stop codon regions for input transcripts. This is an example of customPick function.

**Usage**

```
getStopCodon(trans_ids, txdb, ...)
```

**Arguments**

trans_ids	A character object containing transcript ids.
txdb	A TxDb object.
...	Any additional parameters needed.

**Value**

A numeric object.

## Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
RS2 <- getStopCodon(trans.ids, txdb)
```

---

getTransInfo	<i>Get transcript information</i>
--------------	-----------------------------------

---

## Description

Generate a data frame object that contains information about input genomic feature set and its mapping results over the transcriptome.

## Usage

```
getTransInfo(A, txdb)
```

## Arguments

A	Genomic feature set, which should be a GRanges object.
txdb	A TxDb object.

## Value

A data.frame object containing the following components:

- `index_trans`: The label of transcripts.
- `index_features`: The label of genomic features.
- `seqnames`: The chr name.
- `features_pos`: The starting coordinate of each genomic feature.
- `width_features`: The width of each genomic feature.
- `strand`: The strand type of each genomic feature.
- `trans_ID`: The ids of the transcripts that each feature can be mapped to.

## Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
getTransInfo(A = m6A_sites_data[1:100], txdb)
```

---

GRanges2GRangesList     *Convert a GRanges object to a GRangesList object*

---

## Description

Convert a GRanges object to a GRangesList object. The output region set follows the format required by the main permutation test functions.

## Usage

```
GRanges2GRangesList(A = NULL)
```

## Arguments

A                    A GRanges object.

## Details

If input GRanges object has a metadata named as "group", ranges having the same group number represent a region. If not, a range is a region. A region in the input set will be outputted as a list element IN returned GRangesList object.

## Value

A GRangesList object.

## See Also

[GRanges2GRangesList](#)

## Examples

```
library(GenomicRanges)
GRanges.object <- GRanges(
  Rle(c("chr2", "chr2", "chr1", "chr3")),
  IRanges(1:4, width = 5)
)
# Assign the first and the second ranges to the same element.
GRanges.object$group <- c(1, 1, 2, 3)
GRangesList.object <- GRanges2GRangesList(GRanges.object)
```

---

GRangesList2GRanges     *Convert a GRangesList object to a GRanges object*

---

### Description

Convert a GRangesList object to a GRanges object. The output region set follows the format required by the RgnTX permutation test functions, which should have metadata columns 'group' and 'transcriptsHits'.

### Usage

```
GRangesList2GRanges(A = NULL)
```

### Arguments

A                    A GRangesList object.

### Value

A GRanges object. Its transcript ids (if available) should be contained in a metadata column named "transcriptsHits", which are provided by the names of input GRangesList object.

### See Also

[GRanges2GRangesList](#)

### Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
RS1 <- randomizeTx(txdb, trans.ids, random_num = 100, random_length = 100)

RS1 <- GRangesList2GRanges(RS1)
```

---

overlapCountsTx     *Evaluation function*

---

### Description

This function receives two region sets and returns the number of their overlaps.

### Usage

```
overlapCountsTx(A, B, count_once = TRUE, over_trans = TRUE, ...)
```

**Arguments**

A	Region set 1. A GRangesList object.
B	Region set 2. A GRangesList object.
count_once	Whether the overlap of multiple B regions with a single A region should be counted once or multiple times.
over_trans	Whether the overlapping is counted over the transcriptome or over the genome.
...	Any additional parameters needed.

**Value**

A numeric object.

**See Also**

[overlapCountsTx](#)

**Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
exons.tx0 <- exonsBy(txdb)
regions.A <- exons.tx0[trans.ids]
A <- randomizeTransByOrder(regions.A, random_length = 200)
B <- randomizeTransByOrder(regions.A, random_length = 200)

overlapCountsTx(A, B)
```

---

overlapCountsTxIA      *Evaluation function*

---

**Description**

Evaluation function. This function receives a feature set (with isoform ambiguity) and a transcriptome region set (without isoform ambiguity), and returns a weighted number of overlaps between them.

**Usage**

```
overlapCountsTxIA(A, B, ...)
```

**Arguments**

A	A feature set, which should be GRanges.
B	A region set, which should be GRangesList.
...	Any additional parameters needed.

**Value**

A numeric object.

**See Also**

[overlapWidthTx](#), [distanceTx](#), [overlapCountsTx](#)

**Examples**

```
library(Txdb.Hsapiens.UCSC.hg19.knownGene)
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
txdb <- Txdb.Hsapiens.UCSC.hg19.knownGene
RS1 <- m6A_sites_data[1:100]

trans.info <- getTransInfo(RS1, txdb)
trans.ids <- trans.info[, "trans_ID"]

RS2 <- getStopCodon(trans.ids, txdb = txdb)

# Evaluation step.
orig.ev <- overlapCountsTxIA(RS1, RS2)
```

---

overlapWidthTx

*Evaluation function*

---

**Description**

Evaluation function. This function returns the sum of widths of each overlap between two region sets, i.e., the total number of overlapping nucleotides between two input region sets.

**Usage**

```
overlapWidthTx(A, B, ...)
```

**Arguments**

A	Region set 1. A Granges or GRangesList object.
B	Region set 2. A Granges or GRangesList object.
...	Any additional parameters needed.

**Value**

A numeric object.

**See Also**

[overlapCountsTx](#), [distanceTx](#)

**Examples**

```

library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
A <- randomizeTx(
  txdb, trans.ids, random_num = 20,
  random.length = 100
)
B <- randomizeTx(
  txdb, trans.ids = trans.ids, random_num = 20,
  random_length = 100
)

overlapWidthTx(A, B)

```

---

permTestTx	<i>Perform permutation test</i>
------------	---------------------------------

---

**Description**

Perform permutation test for evaluating spatial association between a feature set and a region set.

**Usage**

```

permTestTx(RS1 = NULL, RS2 = NULL, txdb = NULL, type = "mature",
ntimes = 50, ev_function_1 = overlapCountsTx, ev_function_2 = overlapCountsTx,
pval_z = FALSE, ...)

```

**Arguments**

RS1	The region set to be randomized. It should be in the GRanges or GRangesList format.
RS2	The region set to be compared with. It should be in the GRanges or GRangesList format.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
ntimes	Randomization times.
ev_function_1	Evaluation function defines what statistic to be tested between RS1 and RS2. Default is overlapCountsTx.
ev_function_2	Evaluation function defines what statistic to be tested between each element in RSL and RS2. Default is overlapCountsTx.
pval_z	Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, the p-value is calculated based on a z-test.
...	Any additional parameters needed.

**Details**

permTestTxIA only needs users to input two region sets. It will automatically randomize the first region set into transcriptome.

**Value**

A list object, which is defined to be permTestTx.results class. It contains the following items:

- RSL: Randomized region sets of RS1.
- RS1: The feature set to be randomized.
- RS2: The region set to be compared with the feature set.
- orig.ev: The value of overlapping counts between RS1 and RS2.
- rand.ev: The values of overlapping counts between each element in RSL and RS2.
- pval: p-value of the test.
- zscore: Standard score of the test.

**See Also**

[plotPermResults](#)

**Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
exons.tx0 <- exonsBy(txdb)
trans.ids <- sample(names(exons.tx0), 500)

A <- randomizeTx(txdb, trans.ids, random_num = 100, random_length = 100)
B <- c(randomizeTx(txdb, trans.ids, random_num = 75, random_length = 100), A[1:25])

permTestTx_results <- permTestTx(A, B, txdb, ntimes = 5)
```

---

permTestTxIA

*Perform permutation test*

---

**Description**

Perform permutation test for evaluating spatial association between some features (with isoform ambiguity) and a region set. It randomizes the features and compares it with the region set to see if there is an association between the features and the region set. The difference between this function and [permTestTx](#) is that it is for RNA-related genomic features that have isoform ambiguity, i.e., features that one does not know which transcript they comes from.



**Usage**

```
permTestTxIA(RS1 = NULL,
             RS2 = NULL,
             txdb = NULL,
             type = 'mature',
             ntimes = 50,
             ev_function_1 = overlapCountsTx,
             ev_function_2 = overlapCountsTx,
             pval_z = FALSE,
             ...)
```

**Arguments**

RS1	The feature set to be randomized. It should be in the GRanges or GRangesList format.
RS2	The region set to be compared with. It should be in the GRanges or GRangesList format.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
ntimes	Randomization times.
ev_function_1	Evaluation function defines what statistic to be tested between RS1 and RS2. Default is overlapCountsTx.
ev_function_2	Evaluation function defines what statistic to be tested between each element in RSL and RS2. Default is overlapCountsTx.
pval_z	Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, the p-value is calculated based on a z-test.
...	Any additional parameters needed.

**Details**

permTestTxIA only needs users to input two region sets. It will automatically randomize the first region set into transcriptome.

**Value**

A list object, which is defined to be permTestTx.results class. It contains the following items:

- RSL: Randomized region sets of RS1.
- RS1: The feature set to be randomized.
- RS2: The region set to be compared with the feature set.
- orig.ev: The value of overlapping counts between RS1 and RS2.
- rand.ev: The values of overlapping counts between each element in RSL and RS2.
- pval: p-value of the test.
- zscore: Standard score of the test.

**See Also**[plotPermResults](#)**Examples**

```

library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
RS1 <- m6A_sites_data[1:500]
trans.ids <- getTransInfo(RS1, txdb)[, "trans_ID"]
RS2 <- getStopCodon(trans.ids, txdb)

permTestTx_results <- permTestTxIA(RS1 = RS1, RS2 = RS2,
                                   txdb = txdb, ntimes = 5)

```

---

permTestTxIA\_customPick

*Perform permutation test*


---

**Description**

Perform permutation test for evaluating spatial association between RNA features and a specified kind of regions. The latter is defined by the `customPick_function` argument input by users. The difference between this function and [permTestTx\\_customPick](#) is that it is for RNA-related genomic features that have isoform ambiguity, i.e., features that one does not know which transcript they comes from.

**Usage**

```

permTestTxIA_customPick(RS1 = NULL, txdb = NULL, type = 'mature',
                        customPick_function = NULL, ntimes = 50,
                        ev_function_1 = overlapCountsTxIA, ev_function_2 = overlapCountsTx, pval_z = FALSE, ...)

```

**Arguments**

RS1	The region set to be randomized. It should be in the GRanges or GRangesList format.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
customPick_function	A custom function needs to be inputted by users. The customPick function should have two arguments: a TxDb object and a character object of transcript ids. It returns a specified region over each transcript.



```

type = 'mature',
customPick_function = getStopCodon,
ntimes = 5)

```

---

`permTestTx_customAll` *Perform permutation test*

---

### Description

Perform permutation test for evaluating spatial association between region sets. This permutation test function receives two region sets and a set of randomized region sets of one of them. It evaluates if there is an association between these two region sets.

### Usage

```

permTestTx_customAll(RSL = NULL, RS1 = NULL, RS2 = NULL,
ev_function_1 = overlapCountsTx, ev_function_2 = overlapCountsTx, pval_z = FALSE, ...)

```

### Arguments

RSL	Randomized region sets of RS1. It should be a list object and each element should be in the GRanges or GRangesList format.
RS1	The region set. It should be in the GRanges or GRangesList format.
RS2	The region set to be compared with. It should be in the GRanges or GRangesList format.
ev_function_1	Evaluation function defines what statistic to be tested between RS1 and RS2. Default is <code>overlapCountsTx</code> .
ev_function_2	Evaluation function defines what statistic to be tested between each element in RSL and RS2. Default is <code>overlapCountsTx</code> .
pval_z	Boolean. Default is FALSE. If FALSE, p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, p-value is calculated based on a z-test.
...	Any additional parameters needed.

### Details

`permTestTx_customAll` will use evaluation function `ev_function_1` to calculate the test statistic between RS1 and RS2, and use `ev_function_2` to evaluate the statistic between RSL and RS2. It will also return a p-value and a z-score.

**Value**

A list object, which is defined to be permTestTx.results class. It contains the following items:

- RSL: Randomized region sets of RS1.
- RS1: The feature set to be randomized.
- RS2: The region set to be compared with the feature set.
- orig.ev: The value of overlapping counts between RS1 and RS2.
- rand.ev: The values of overlapping counts between each element in RSL and RS2.
- pval: p-value of the test.
- zscore: Standard score of the test.

**Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids1<- c("170")
RS1 <- randomizeTx(txdb = txdb, trans_ids = trans.ids1,
                  random_num = 20, random_length = 100)
RS2 <- randomizeTx(txdb = txdb, trans_ids = trans.ids1,
                  random_num = 20, random_length = 100)
trans.ids2 <- c("170", "782", "974", "1364", "1387")
RSL <- randomizeTx(txdb = txdb, trans_ids = trans.ids2,
                  random_num = 20, random_length = 100, N = 10)
permTestTx_results <- permTestTx_customAll(RSL = RSL, RS1 = RS1, RS2 = RS2)
```

---

permTestTx\_customPick *Perform permutation test*

---

**Description**

Perform permutation test for evaluating spatial association between a feature set and the customPick regions. The latter is defined by the customPick\_function argument provided by users.

**Usage**

```
permTestTx_customPick(RS1 = NULL, txdb = NULL, type = "mature",
                     customPick_function = NULL, ntimes = 50, ev_function_1 = overlapCountsTx,
                     ev_function_2 = overlapCountsTx, pval_z = FALSE, ...)
```

**Arguments**

RS1	The feature set to be randomized. It should be in the GRanges or GRangesList format.
txdb	A TxDb object.

<code>type</code>	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
<code>customPick_function</code>	A custom function needs to be inputted by users. The custom function should have two arguments: a TxDb object and a character object of transcript ids. It returns a part of region of each transcript.
<code>ntimes</code>	Randomization times.
<code>ev_function_1</code>	Evaluation function defines what statistic to be tested between RS1 and RS2. Default is <code>overlapCountsTx</code> .
<code>ev_function_2</code>	Evaluation function defines what statistic to be tested between each element in RSL and RS2. Default is <code>overlapCountsTx</code> .
<code>pval_z</code>	Boolean. Default is FALSE. If FALSE, the p-value is calculated based on the number of random evaluations is larger or less than the initial evaluation. If TRUE, the p-value is calculated based on a z-test.
<code>...</code>	Any additional parameters needed.

### Details

Each feature in RS1 is only mapped with the customPick regions over its transcript (picked by the customPick\_function). The output `orig.ev` is the number of features that have overlap with its customPick region. The set of randomized region sets is outputted as RSL. The overlapping counts between each set in RSL with RS2 is outputted as `rand.ev`.

### Value

A list object, which is defined to be `permTestTx.results` class. It contains the following items:

- `RSL`: Randomized region sets of RS1.
- `RS1`: The feature set to be randomized.
- `RS2`: The region set to be compared with the feature set.
- `orig.ev`: The value of overlapping counts between RS1 and RS2.
- `rand.ev`: The values of overlapping counts between each element in RSL and RS2.
- `pval`: p-value of the test.
- `zscore`: Standard score of the test.

### See Also

[plotPermResults](#)

### Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
exons.tx0 <- exonsBy(txdb)
trans.ids <- sample(names(exons.tx0), 100)
```

```

RS1 <- randomizeTx(txdb, trans.ids, random_num = 100,
random_length = 200, type = 'CDS')
getCDS = function(txdb, trans.id){
  cds.tx0 <- cdsBy(txdb, use.names=FALSE)
  cds.names <- as.character(intersect(names(cds.tx0), trans.id))
  cds = cds.tx0[cds.names]
  return(cds)
}

permTestTx_results <- permTestTx_customPick(RS1,txdb,
customPick_function = getCDS, ntimes = 5)

```

---

plotPermResults      *Plot permutation test results*

---

### Description

Show a graphical representation of permutation test.

### Usage

```
plotPermResults(permTestTx_results = NULL, breaks = 15, alpha = 0.05,
test_type = "one-sided", binwidth = NULL)
```

### Arguments

permTestTx_results	A permTestTx.results list object, which can be generated by the permTestTx function.
breaks	Histogram breaks. Default is 15.
alpha	Significance level.
test_type	The type of the test. This argument only receives either two options "one-sided" or "two-sided". Default is "one-sided".
binwidth	Histogram binwidth.

### Value

A plot object.

### See Also

[permTestTx](#)

### Examples

```

file <- system.file(package="RgnTX", "extdata", "permTestTx_results.rds")
permTestTx_results <- readRDS(file)
p_a <- plotPermResults(permTestTx_results, binwidth = 1)
p_a

```

plotShiftedZScoreTx *Plot shifted z scores*

---

**Description**

Plot shifted z scores for permutation test results.

**Usage**

```
plotShiftedZScoreTx(shitedZScoresTx_results)
```

**Arguments**

shitedZScoresTx\_results  
A shitedZScoreTx.results object.

**Value**

A plot.

**See Also**

[shiftedZScoreTx](#)

**Examples**

```
file <- system.file(package="RgnTX", "extdata", "shiftedZScoreTx_results1.rds")
shiftedZScoreTx_results <- readRDS(file)
p1 <- plotShiftedZScoreTx(shiftedZScoreTx_results)
p1
```

---

randomizeFeaturesTx *Randomize features into transcriptome*

---

**Description**

Randomize features into transcriptome.

**Usage**

```
randomizeFeaturesTx(RS, txdb, type = "mature", N = 1, ...)
```



**Arguments**

RS	The feature to be randomized. It should be a GRanges or GRangesList object.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
N	The number of iterations.
...	Any additional parameters needed.

**Value**

A GRangesList object. The name of each element is the id of the transcript where the corresponding range is located.

**See Also**

[randomizeTransByOrder](#), [randomizeFeaturesTxIA](#), [randomizeTx](#)

**Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
RS1 <- randomizeTx(txdb, trans.ids, random_num = 100, random_length = 100)
RS <- randomizeFeaturesTx(RS1, txdb, N = 1)
```

---

randomizeFeaturesTxIA *Randomize features into transcriptome*

---

**Description**

Randomize features into transcriptome, especially for the features that have isoform ambiguity.

**Usage**

```
randomizeFeaturesTxIA(RS, txdb, type = "mature", N = 1, ...)
```

**Arguments**

RS	The feature being randomized. It should be a GRanges or GRangesList object.
txdb	A TxDb object.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
N	Randomization times.
...	Any additional parameters needed.

**Value**

A GRangesList object. The name of each element is the id of the transcript where the corresponding range is located.

**See Also**

[randomizeTransByOrder](#), [randomizeFeaturesTx](#), [randomizeTx](#)

**Examples**

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
RS1 <- m6A_sites_data[1:100]
RS <- randomizeFeaturesTxIA(RS1, txdb, N = 1)
```

---

randomizeTransByOrder *Randomize features into transcriptome*

---

**Description**

This function receives a GRangesList object and picks a random region within each list element of this object. The length of the region to be picked is decided by the input random\_length argument.

**Usage**

```
randomizeTransByOrder(regions_A, random_length = 20)
```

**Arguments**

regions\_A      A GRangesList object. The name of each list element should be the corresponding transcript id.

random\_length    A numeric object.

**Value**

A GRangesList object. The name of each list element should be the corresponding transcript id.

**See Also**

[randomizeTx](#), [randomizeFeaturesTx](#), [randomizeFeaturesTxIA](#)

## Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
exons.tx0 <- exonsBy(txdb)
trans.ids <- sample(names(exons.tx0), 500)
regions.A <- exons.tx0[trans.ids]
RS <- randomizeTransByOrder(regions.A, random_length = 20)
```

---

randomizeTx

*Get randomized regions over transcriptome*

---

## Description

Pick random regions over specified transcripts.

## Usage

```
randomizeTx(txdb, trans_ids = 'all',
            random_num = 100, random_length = 20, type = 'mature', N = 1, ...)
```

## Arguments

txdb	A TxDb object.
trans_ids	The ids of transcripts, which should be a character object. Random regions will be picked from these transcripts. If this argument takes the default value 'all', the scope of picking random regions will be the whole transcriptome.
random_num	The number of regions to be picked.
random_length	The length of regions to be picked.
type	A character object. Default is "mature". It accepts options "mature", "full", "fiveUTR", "CDS" or "threeUTR", with which one can get corresponding types of transcriptome regions.
N	Randomization times.
...	Any additional parameters needed.

## Value

A GRangesList object. The name of each element is the id of the transcript where the corresponding range is located.

## See Also

[randomizeTransByOrder](#), [randomizeFeaturesTx](#), [randomizeFeaturesTxIA](#)

## Examples

```
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
trans.ids <- c("170", "782", "974", "1364", "1387")
RS1 <- randomizeTx(txdb, trans.ids, random_num = 100, random_length = 100)
```

---

shiftedZScoreTx	<i>Calculate shifted z scores</i>
-----------------	-----------------------------------

---

## Description

Calculate shifted z scores for permutation test results.

## Usage

```
shiftedZScoreTx(permTestTx_results = NULL, txdb = NULL,
window = 200, step = 20, ev_function_1 = overlapCountsTx, ...)
```

## Arguments

permTestTx_results	A permTestTx.results object.
txdb	A TxDb object.
window	The window of the whole shifting.
step	The step of each shifting.
ev_function_1	Evaluation function. Default is overlapCountsTx.
...	Any additional parameters needed.

## Details

see examples in [plotShiftedZScoreTx](#)

## Value

A list object, which is defined to be shiftedZScore.results class. It contains the following items:

- shifted.z.scores: Standard z-scores after shifting.
- window: Window of the whole shifting.
- step: Step of each shifting.
- original.z.score: Original standard score.

## See Also

[plotShiftedZScoreTx](#)

**Examples**

```

library(TxDb.Hsapiens.UCSC.hg19.knownGene)
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene
file <- system.file(package="RgnTX", "extdata/m6A_sites_data.rds")
m6A_sites_data <- readRDS(file)
RS1 <- m6A_sites_data[1:500]
permTestTx_results <- permTestTxIA_customPick(RS1 = RS1,
                                              txdb = txdb,
                                              customPick_function = getStopCodon,
                                              ntimes = 5)
shiftedZScoreTx_results <- shiftedZScoreTx(permTestTx_results, txdb = txdb,
                                           window = 2000,
                                           step = 200,
                                           ev_function_1 = overlapCountsTxIA)

```

---

shiftTx	<i>Shift over transcripts</i>
---------	-------------------------------

---

**Description**

Calculate positional shifting over transcript regions. This function accepts a feature set and outputs a region set from it. Each output region is from each input feature.

**Usage**

```
shiftTx(regions, start, width, direction, strand)
```

**Arguments**

regions	A feature set following the format indicated in vignette section 3. Either to be GRanges or GRangesList.
start	Starting positions. Each value represents a starting position in each input feature.
width	Widths. Each value represents a width of each region to be picked from each feature.
direction	Either to be character "left" or "right", which means the direction to which the starting position is shifting. The former means moving to the direction of 5' while the latter means moving to 3'.
strand	The strand type of the transcripts. It receives "+" or "-".

**Value**

A Granges object.

**Examples**

```

# Take five transcripts.
# Extract the last 200 nt regions from their CDS part.
library(TxDb.Hsapiens.UCSC.hg19.knownGene)
trans.id.pstv <- c("170", "782", "974", "1364", "1387")
txdb <- TxDb.Hsapiens.UCSC.hg19.knownGene

# download the CDS part of all transcriptome
cds.tx0 <- cdsBy(txdb, use.names = FALSE)

# pick the CDS part of these five transcripts
cds.p <- cds.tx0[trans.id.pstv]

width <- 200
start <- as.numeric(max(end(cds.p)))
R.cds.last200 <- shiftTx(cds.p, start = start, width = width, direction = 'left', strand = "+")

```

---

vector2GRangesList      *vector2GRangesList*

---

**Description**

Generate GRangesList object from vectors. The output region set follows the format required by the main permutation test functions.

**Usage**

```
vector2GRangesList(RefSeqID, targetName, strand, blockSizes, targetStart)
```

**Arguments**

RefSeqID	The name of each element.
targetName	The seqnames of each range.
strand	The strand of each range.
blockSizes	The width of each range.
targetStart	The start coordinate of each range.

**Value**

A GRangesList object.

**See Also**

[GRanges2GRangesList](#)

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